

Technical Information

Phenol Red Salicin Broth

Product Code: DM 2011

Application: - Phenol Red Salicin Broth is used for salicin fermentation studies of microorganisms.

Composition**

| Ingredients | Gms / Litre |
|---------------------|-------------|
| Proteose peptone | 10.000 |
| Beef extract | 1.000 |
| Sodium chloride | 5.000 |
| Salicin | 5.000 |
| Phenol red | 0.018 |
| Final pH (at 25°C) | 7.4±0.2 |

**Formula adjusted, standardized to suit performance parameters

Principle & Interpretation

Phenol Red Broth Medium formulated by Vera ⁽²⁾ is recommended to determine the fermentation behavior of different carbohydrates for the differentiation of microorganisms ⁽³⁻⁵⁾. Phenol Red Broth Medium with various carbohydrates serves as a differential medium by helping in differentiation of various species and genera by their ability to ferment the specific carbohydrate, with the production of acid or acid and gas ⁽⁶⁾.

Phenol Red Salicin Broth is used to study salicin fermentation in various bacteria.

Proteose peptone and beef extract serve as sources for carbon and nitrogen. Sodium chloride is the osmotic stabilizer. Phenol red is the pH indicator, which turns yellow at acidic pH i.e. on fermentation of salicin. Gas formation is seen in Durhams tubes. All the members of *Enterobacteriaceae* family grow well in this medium. In addition to producing a pH colour shift, the production of mixed acids, notably butyric acids, often results in a pungent, foul odour from the culture medium ⁽¹⁾.

Methodology

Suspend 21 grams of powder media in 1000 ml distilled water and mix well. Shake well & heat if necessary to ensure complete dissolution. Distribute in fermentation tubes (tubes containing inverted Durham's tubes). Sterilize by autoclaving at 15 lbs pressure (121°C) for 15 minutes.

Quality Control

Physical Appearance

Light yellow to pink coloured homogeneous free flowing powder

Colour and Clarity of prepared medium

Red coloured clear solution without any precipitate

Reaction

Reaction of 2.1% w/v aqueous solution at 25°C. pH : 7.4±0.2

pH Range 7.20-7.60

Cultural Response/ characteristics

DM 2011: Cultural characteristics observed after an incubation at 35 - 37°C for 18 - 24 hours.



Dehydrated Culture Media
Bases / Media Supplements

| Organism | Inoculum (CFU) | Growth | Acid | Gas |
|-----------------------------------|----------------|-----------|-------------------------------------|-------------------|
| Citrobacter freundii ATCC 8090 | 50-100 | luxuriant | Negative reaction, no colour change | Negative reaction |
| Escherichia coli ATCC 25922 | 50-100 | luxuriant | Positive reaction, yellow colour | Positive reaction |
| Enterobacter aerogenes ATCC 13048 | 50-100 | luxuriant | Positive reaction, yellow colour | Positive reaction |
| Klebsiella pneumoniae ATCC 13883 | 50-100 | luxuriant | Positive reaction, yellow colour | Positive reaction |
| Proteus vulgaris ATCC 13315 | 50-100 | luxuriant | Positive reaction, yellow colour | Positive reaction |
| Salmonella Typhi ATCC 6539 | 50-100 | luxuriant | Negative reaction, no colour change | Negative reaction |
| Salmonella Typhimurium ATCC 14028 | 50-100 | luxuriant | Negative reaction, no colour change | Negative reaction |
| Serratia marcescens ATCC 8100 | 50-100 | luxuriant | Positive reaction, yellow colour | Weak reaction |
| Shigella flexneri ATCC 12022 | 50-100 | luxuriant | Negative reaction, no colour change | Negative reaction |

Storage and Shelf Life

Dried Media: Store below 30°C in tightly closed container and use before expiry date as mentioned on the label.

Prepared Media: 2-8° in sealable plastic bags for 2-5 days.

Further Reading

1. Koneman E. W., Allen S. D., Janda W.M., Schreckenberger P.C., Winn W.C. Jr., 1992, Colour Atlas and Textbook of Diagnostic Microbiology, 4th Ed., J. B. Lippincott Company
2. Vera H. D., 1950, Am. J. Public Health, 40, 1267
3. MacFaddin J. F., 1985, Media for Isolation-Cultivation-Identification -Maintenance of Medical Bacteria, Vol. I, Williams and Wilkins, Baltimore.
4. Finegold S. M. and Baron E. J., 1986, Bailey and Scotts Diagnostic Microbiology, 7th Ed., The C.V. Mosby Co., St. Louis.
5. Ewing W. H., 1986, Edwards and Ewings Identification of Enterobacteriaceae, 4th ed., Elsevier Science Publishing Co., Inc., New York.
6. MacFaddin J. F., 2000, Biochemical tests for Identification of Medical Bacteria, 3rd ed., Lippincott, Williams and Wilkins, Baltimore.

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