

Technical Information

TCBS Agar (Selective)

Product Code: DM 1870

Application: - TCBS Agar is recommended for the selective isolation of *Vibrio cholerae* and other enteropathogenic *Vibrios*.

Composition**

Ingredients	Gms / Litre
Peptone, special	10.000
Yeast extract	5.000
Sodium citrate	10.000
Sodium thiosulphate	10.000
Sodium cholate	3.000
Oxgall	5.000
Sucrose	20.000
Sodium chloride	10.000
Ferric citrate	1.000
Bromo thymol blue	0.040
Thymol blue	0.040
Agar	15.000
Final pH (at 25°C)	8.8±0.2

**Formula adjusted, standardized to suit performance parameters

Principle & Interpretation

TCBS Agar was developed by Kobayashi et al ⁽¹⁾, who modified the selective medium of Nakanishi ⁽²⁾. Although this medium was originally designed for the isolation of *V. cholerae* and *V. parahaemolyticus*, most *Vibrios* grow as large colonies with many different colonial morphologies. TCBS Agar is also recommended by APHA for the selective isolation of *V. cholerae* and *V. parahaemolyticus* ^(3, 4). Enrichment in Alkaline Peptone Water (DM1618), followed by isolation on TCBS Agar is routinely used for isolation of *V. cholerae* ⁽⁵⁻⁷⁾. TCBS Agar, Selective has an additional selective ingredient i.e. sodium cholate for improved selectivity.

Peptone special and yeast extract provide nitrogenous compounds, vitamin B complex and other essential growth nutrients. Oxgall, a derivative of bile salts and sodium citrate inhibit growth of gram-positive bacteria and coliforms ⁽⁸⁾. Sodium thiosulphate acts as a good source of sulphur, which in combination with ferric citrate detects the production of hydrogen sulphide. For the metabolism of *Vibrios*, sucrose is added as a fermentable carbohydrate. *Vibrio* that is able to utilize sucrose will form yellow colonies. Bromothymol blue and thymol blue are the pH indicators. The alkaline pH of the medium improves the recovery of *V. cholerae*. *V. alginolyticus* also produce yellow colonies. *V. parahaemolyticus* is a sucrose non-fermenting organism and therefore produces blue-green colonies, as does *V. vulnificus*. *Proteus* species that are sucrose-fermenters may form yellow colonies ⁽⁹⁾. TCBS Agar is not a suitable medium for oxidase testing of *Vibrio* species ⁽¹⁰⁾. A few strains of *V. cholerae* may appear green or colourless on TCBS Agar due to delayed sucrose fermentation ⁽⁹⁾.

TCBS Agar is highly selective for *Vibrio* species. However, occasional isolates of *Pseudomonas* and *Aeromonas* may also form blue green colonies on TCBS Agar ⁽⁹⁾. Any H₂S negative colony of TCBS Agar can be considered presumptive positive for *Vibrio*.

The medium should be inoculated heavily with faecal specimens because growth of few species may be inhibited on the medium due to fermentation of sucrose and accumulation of acids.

Methodology

Suspend 89.08 grams of powder media in 1000 ml distilled water. Shake well & heat to dissolve the medium completely. DO NOT AUTOCLAVE. Cool to 50°C and pour into sterile Petri plates.

Quality Control

Physical Appearance

Light yellow to light tan homogeneous free flowing powder

Gelling

Firm, comparable with 1.5% Agar gel

Colour and Clarity of prepared medium

Bluish green coloured clear to slightly opalescent gel forms in Petri plates

Reaction

Reaction of 8.9% w/v aqueous solution at 25°C. pH : 8.8±0.2

pH range

8.60-9.00

Cultural Response/Characteristics

DM 1870: Cultural characteristics observed after an incubation at 35-37°C for 18-24 hours.

Organism	Inoculum (CFU)	Growth	Recovery	Colour of colony
<i>Escherichia coli</i> ATCC 25922	≥10 ³	inhibited	0%	
<i>Shigella flexneri</i> ATCC 12022	≥10 ³	inhibited	0%	
<i>Enterococcus faecalis</i> ATCC29212	≥10 ³	inhibited	0%	
<i>Vibrio cholerae</i> ATCC 15748	50-100	good- luxuriant	≥50%	yellow
<i>Vibrio fluvialis</i> ATCC 33809	50-100	good- luxuriant	≥50%	yellow
<i>Vibrio parahaemolyticus</i> ATCC 17802	50-100	good- luxuriant	≥50%	bluish green
<i>Vibrio vulnificus</i> ATCC 29306	50-100	fair-good	30-40%	greenish yellow

Storage and Shelf Life

Dried Media: Store below 30°C in tightly closed container and use before expiry date as mentioned on the label.

Prepared Media: 2-8⁰ in sealable plastic bags for 2-5 days.

Further Reading

1. Kobayashi T., Enomoto S., Sakazaki R., and Kuwahara S., 1963, Jap. J. Bacteriol., 18: 387.
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3. Downes F. P. and Ito K., (Eds.), 2001, Compendium of Methods for the Microbiological Examination of Foods, 4th Ed., American Public Health Association, Washington, D.C.
4. Clesceri L. S., Greenberg A. E. and Eaton A. D., (Eds.), 1998, Standard Methods for the Examination of Water and Wastewater, 20th Ed., American Public Health Association, Washington, D.C.
5. Murray P. R., Baron J. H., Pfaller M. A., Jorgensen J. H. and Tenover F. C., (Eds.), 2003, Manual of Clinical Microbiology, 8th Ed., American Society for Microbiology, Washington, D.C.
6. Furniss A. L., Lee J. V. and Donovan T. J., 1978, The Vibrios, Public Health Laboratory Service Monograph Series No. 11, Maidstone Public Health Laboratory, H.M.S.O., London, England.
7. Forbes B. A., Sahm A. S. and Weissfeld D. F., 1998, Bailey & Scotts Diagnostic Microbiology, 10th Ed., Mosby, Inc. St. Louis, Mo.
8. Howard B., 1994, Clinical and Pathogenic Microbiology, 2nd Ed., The C.V. Mosby.
9. MacFaddin J. F., 1985, Media for Isolation-Cultivation-Identification-Maintenance of Medical Bacteria, Vol. 1, Williams & Wilkins, Baltimore, Md.
10. Morris G. K., Merson M. H., Huq A. K., Kibrya A. K. and Black R., 1979, J. Clin. Microbiol., 9:79

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