

Technical Information

Pseudomonas Agar Medium for Detection of Pyocyanin

Product Code: DM 1119U

Application: - Pseudomonas Agar Medium for detection of Pyocyanin is used for the detection of pyocyanin production by *Pseudomonas* species in accordance with United States Pharmacopoeia.

Composition**

Ingredients	Gms / Litre
Pancreatic digest of gelatin	20.000
Anhydrous potassium sulphate	10.000
Anhydrous magnesium chloride	1.400
Agar	15.000
pH after sterilization (at 25°C)	7.2±0.2

**Formula adjusted, standardized to suit performance parameters

Principle & Interpretation

Pseudomonas Agar Medium for detection of Pyocyanin is based on the formulation described by King et al (1) and as used by U.S. Pharmacopoeia (2) for detecting pyocyanin, a water soluble pigment by *Pseudomonas* species from clinical specimens such as stools, wounds, and urine. (3). It is also recommended for microbial limit tests for pharmaceutical and other biological preparations by USP. *Pseudomonas* species are commonly isolated pathogen and is the significant causative agent of nosocomial, skin and burn infections. *Pseudomonas* strains are reported to produce phenazine pigments like Pyocyanin- blue green redox-active secondary metabolite pigment, pyorubin-rust brown pigment, -oxyphenazine- a breakdown product of Pyocyanin, pyoverdin-a water soluble yellow green pigments also known as fluorescein. Pyocyanin is readily recovered in large quantities in sputum from patients with cystic fibrosis, an infection caused by *Pseudomonas* (4,5). This medium enhances the formation of Pyocyanin and/or pyorubin and reduces that of fluorescein.

Pancreatic digest of casein supplies essential nutrients for growth of *Pseudomonas*, while glycerol provides carbon and energy to the cell. The pyocyanin pigment diffuses from the colonies of *Pseudomonas* into the agar and shows blue colouration. Potassium sulphate and magnesium chloride enhances the pyocyanin production and suppresses the fluorescein production. Low content of phosphorous in the medium also aids in inhibiting the production of fluorescein. Some *Pseudomonas* strains produce small amounts of fluorescein resulting in a blue-green colouration.

Strains of *Pseudomonas aeruginosa* that may fail to produce pyocyanin are not detected in this medium. Production of other pigments may mask the presence of pyocyanin.

Methodology

Suspend 46.4 grams of dehydrated powder media in 1000 ml distilled water containing 10 ml glycerin. Mix thoroughly & heat to boiling to dissolve the medium completely. Sterilize by autoclaving at 15 lbs pressure (121°C) for 15 minutes or as per validated cycle.

Quality Control

Appearance

Cream to yellow homogeneous free flowing powder.

Gelling

Firm comparable with 1.5% Agar gel.

Colour and Clarity

Yellow coloured clear to slightly opalescent gel forms in Petri plates.

pH Range

7.00-7.40

Cultural Response

Growth Promotion is carried out in accordance with the harmonized method of USP. Cultural response was observed after an incubation at 33-37°C for not less than 3 days. Recovery rate is considered as 100% for bacteria growth on Soyabean Casein Digest Agar.

Cultural Response

Organism	Inoculum (CFU)	Observed Lot value (CFU)	Recovery	Characteristic colonial morphology	Fluorescence in UV light	Oxidase
Test for Pseudomonas aeruginosa						
<i>Pseudomonas aeruginosa</i> ATCC 9027	50 -100	35 -100	>=70 %	Generally greenish	Blue	positive
Additional Microbiological Testing						
<i>Pseudomonas aeruginosa</i> ATCC 27853	50 -100	35 -100	>=70 %	Generally greenish	Blue	positive

Storage and Shelf Life

Dried Media: Store below 30°C in tightly closed container and the prepared medium at 2 - 8°C. Use before expiry date on the label.

Prepared Media: 2-8° in sealable plastic bags for 2-5 days.

Further Reading

1. King, Ward and Raney, 1954, J.Lab. and Clin. Med., 44:301
2. United States Pharmacopoeia, 2008, United States Pharmacopoeia Convention, Inc., Rockville, MD.
3. MacFaddin J., 1985, Media for Isolation-Cultivation-Identification-Maintenance of Medical Bacteria, Vol. I, Williams an Wilkins, Baltimore.
4. Daly J A, Boshard R, and Matsen J M, 1984, J Clin Microbiol. 19: 742
5. Lau GW, Hassett DJ, Ran H, Kong F., 2004. Trends Mol Med. 10:599

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