

## Technical Information

### Feeley Gorman Agar (F.G.Agar)

**Product Code: DM 1811**

**Application:** - Feeley Gorman Agar is used for the isolation and presumptive identification of *Legionella* species.

#### Composition\*\*

Ingredients	Gms / Litre
Casein acid hydrolysate	17.500
Beef extract	3.000
Starch	1.500
L-Cysteine hydrochloride	0.400
Ferric pyrophosphate, soluble	0.250
Agar	17.000
Final pH ( at 25°C)	6.9±0.2

\*\*Formula adjusted, standardized to suit performance parameters

#### Principle & Interpretation

Feeley et al formulated (1, 2) Feeley Gorman Agar, which is used as nonselective enrichment medium for isolation of *Legionella* species. *Legionella* is a gram-negative bacterium, including species that cause legionellosis or Legionnaires' disease, most notably *L. pneumophila*(3). *Legionella* species are the causative agent of the human Legionnaires' disease and the lesser form, Pontiac fever. *Legionella* transmission occurs via aerosols- inhalation of mist droplets containing the bacteria. Person-to-person transmission of *Legionella* has not been demonstrated (4).

*Legionella* are nutritionally fastidious and require L-cysteine and iron salts for their growth, which are provided in the medium.

*Legionella* species are highly pathogenic microorganisms. Certain safety precautions must be taken when handling *Legionella* cultures.

Casein acid hydrolysate, beef extract, L-cysteine hydrochloride and ferric pyrophosphate serve as sources of nutrients. Incubation should be carried out in the presence of 2.5% carbon dioxide but if it exceeds the limit, *Legionella* growth is prevented due to formation of acidic condition. It is used to inoculate F.G. Agar and Legionella Agar (DM1809) with supplements simultaneously, as *Legionella* usually do not grow initially on F.G. agar. *Legionella* species can be identified by their characteristic fluorescence in presence of UV light (5, 6).

Safety Precautions for handling specimens and cultures. Use bacteriological safety hood (Biosafety cabinet). Wear gown, mask and gloves.

Decontaminate work surface with either 5% hypochlorite or 5% phenol. Autoclave all materials before discarding or cleaning. Since

*Legionella* disease is primarily a pulmonary infection, prevention and containment of aerosols is essential (7).

#### Methodology

Suspend 39.65 grams of dehydrated media in 1000 ml distilled water. Mix thoroughly & heat to boiling to dissolve the medium completely. Sterilize by autoclaving at 15 lbs pressure (121°C) for 15 minutes. Cool to 45 - 50°C. Shake well before pour into sterile Petri plates.

#### Quality Control

##### Appearance

Cream to yellow homogeneous free flowing powder

##### Gelling

Firm, comparable with 1.7% agar gel.



Dehydrated Culture Media  
Bases / Media Supplements

#### Colour and Clarity

Yellow coloured, clear to slightly opalescent gel forms in Petri plates

#### Reaction

Reaction of 4.0% w/v aqueous solution at 25°C. pH : 6.9±0.2

#### Ph Range

6.70-7.10

#### Cultural Response

DM1811: Cultural characteristics observed in presence of 2.5% Carbon dioxide (CO<sub>2</sub>) after an incubation at 35-37°C for 4 days .

#### Cultural Response

Organism	Growth	Fluorescence under 366 nm
<i>Legionella bozemannii</i> ATCC 33217	good-luxuriant	blue-white
<i>Legionella micdadei</i> ATCC 33218	good-luxuriant	none
<i>Legionella pneumophila</i> ATCC 33153	good-luxuriant	bright yellow

## Storage and Shelf Life

**Dried Media:** Store below 30°C in tightly closed container and the prepared medium at 2 - 8°C away from light. Use before expiry date on the label.

**Prepared Media:** 2-8° in sealable plastic bags for 2-5 days.

## Further Reading

1. Feeley J. C. et al, 1978, J. Clin. Microbiol., 8(3): 320.
2. Feeley J. C. et al, 1979, J. Clin. Microbiol., 10(4):437.
3. Ryan K. J., Ray C. G. (Eds.), 2004, Sherris Medical Microbiology, 4th Edition, McGraw Hill.
4. Winn, W. C. Jr. ,1996, Legionella (In: Baron's Medical Microbiology, Barron, S. et al, (Eds.), 4th Edition, University of Texas Medical Branch
5. Herbert G. A. et al, 1959, Ann. Intern, Med., 92(1):45.
6. Herbert G. A. et al, 1980, Ann. Intern. Med., 92(1):53.
7. MacFaddin J. F., Vol. I, 1985, Media for Isolation Cultivation-Identification-Maintenance of Medical Bacteria, Williams and Wilkins, Baltimore/ London, pg.307-308.

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