



# **Product Specification**

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# **Technical Information**

## **Trypsin 0.25% Solution 1X** 0.25% Trypsin in Hank's Balanced Salt Solution With Phenol red Without Calcium and Magnesium

### Product Code:TCL1047

### Application:-

Trypsin Molecular Weight: 23.4kDa CAS No: 9002-07-07

EC No: 3.4.21.4

Trypsin is a serine protease derived from porcine pancreas. It is a single chain polypeptide of 223 amino acid residue with substrate specificity based on positively charged Lysine and Arginine side chains. Trypsin predominantly cleaves peptide chains at the carboxyl sides of Lysine and Arginine, except when either is followed by Proline. It is most commonly used for dissociation and disaggregation of adherent cells. TCL1047 is 0.25% Trypsin in Hank's balanced salt solution.

### Activity:

One BAEE unit will produce a ΔA253nm of 0.001 per minute with BAEE as substrate at pH 7.6 at 25°C in a reaction volume of 3.2ml (1cm light path). One TAME unit hydrolyzes 1umole of p-toluenesulfonyl-L-arginine methyl ester (TAME) per minute at 25°C, pH 8.2, in the presence of 0.001M calcium ion. One USP trypsin unit is the activity causing a change inabsorbance of 0.003 per minute under the conditions specified. Activity Conversion: 1 TAME unit = 19.2 USP or NF units = 57.5 BAEE Units

## Methodology

### Dissociation of cells from culture vessel

1. Remove the spent medium from the culture vessel by aspiration.

- 2. Wash the monolayer by adding balanced salt solution without calcium and magnesium to the side of the flask opposite the cells.
- 3. Rinse the cell sheet by rocking the flask for 1 to 2 minutes and discard the wash solution.

4. Add Trypsin or Trypsin EDTA solution to the side of the flask opposite the cells. The volume should be sufficient enough to completely cover the monolayer of the cells.

5. Rock the flask to ensure that the dissociation solution covers the cell sheet.

6. Incubate the flask at 37°C for 2 to 3 minutes. Monitor the process by observing the flask under inverted microscope. When dissociation is complete, the cells will be in suspension and appear rounded. In addition to rocking gently, flasks of cell lines that are characteristically difficult to remove from substratum may be tapped to expedite removal.

Note:- The exact time needed to dissociate cells will vary according to the cell line. The dissociation process should be monitored closely to avoid cell damage.

7. Once the cell dissociation is complete add serum containing complete medium to the flask to inhibit the tryptic activity which may further damage the cells.

8. Disperse the cells into a single cell suspension by pipetting repeatedly.

9. Count and subculture the cells.

#### Note:

1. Concentration of Trypsin or Trypsin EDTA solution used for dissociation should be determined empirically for individual cell lines.

2. Time required for dissociation of cells from surface depends on cell type, cell density, potency of trypsin, serum concentration in growth medium and time since last subculture.

3. For serum free media, use Soybean Trypsin inhibitor (TCL1068) 1:1 to neutralize the action of trypsin.





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## Quality control

#### Appearance

Red, clear solution.

рΗ

7.00 -7.60

### Osmolality in mOsm/Kg H<sub>2</sub>O

270.00 -310.00

### Sterility

No bacterial or fungal growth is observed after 14 days of incubation, as per USP specification

### Cultural Response

Cell Dissociation Test.

### Storage and Shelf Life

Shelf life of the product is 24 months.

Upon receipt store at -20°C in a freezer that is not self defrosting. Once thawed the product is stable about 2 weeks at 2 - 8°C. Repeated freezing and thawing reduces enzymatic activity and should be avoided. Once thawed, the solution can be aliquoted in smaller volumes and frozen for future use. Use before expiry date given on the product label.

### **Disclaimer**:

- User must ensure suitability of the product(s) in their application prior to use.
- The product conforms solely to the technical information provided in this booklet and to the best of knowledge research and development work carried at **CDH** is true and accurate.
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