



Ready Prepared Media

## Technical Information

### Sabouraud Dextrose Agar Plate w/ 1% Glycerol ( $\gamma$ -irradiated)(Triple Pack) ( $\gamma$ -irradiated) (Triple Pack)

**Product Code: PM 1063AGT**

**Application:** Recommended for cultivation of yeasts, moulds and aciduric microorganisms.

#### Composition\*\*

Ingredients	Gms / Litre
Dextrose (Glucose)	40.000
Mycological, peptone	10.000
Agar	15.000
Glycerol	10ml
Final pH ( at 25°C)	5.6±0.2

\*\*Formula adjusted, standardized to suit performance parameters

#### Principle & Interpretation

Sabouraud Dextrose Agar is Carlier's modification (3) of the formulation described by is a modification of Sabouraud Dextrose Agar which is described by Sabouraud (7) for the cultivation of fungi (yeasts, moulds), particularly useful for the fungi associated with skin infections. This medium is also employed to determine microbial contamination in food, cosmetics, and clinical specimens (2).

Mycological Peptone provides nitrogenous compounds. Dextrose provides an energy source. High dextrose concentration and low pH favors fungal growth and inhibits contaminating bacteria from test samples (6).

#### Type of specimen

Food samples; Cosmetics

#### Specimen Collection and Handling

For food and dairy samples, follow appropriate techniques for sample collection and processing as per guidelines (3,4,5). For water samples, follow appropriate techniques for sample collection, processing as per guidelines and local standards (10). For clinical samples follow appropriate techniques for handling specimens as per established guidelines (11,12). After use, contaminated materials must be sterilized by autoclaving before discarding.

#### Warning and Precautions

Read the label before opening the pack. Wear protective gloves/protective clothing/eye protection/face protection. Follow good microbiological lab practices while handling specimens and culture. Standard precautions as per established guidelines should be followed while handling specimens. Safety guidelines may be referred in individual safety data sheets.



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## Limitations

1. Some pathogenic fungi may produce infective spores which are easily dispersed in air, so examination should be carried out in safety cabinet.
2. Individual organisms differ in their growth requirement and may show variable growth patterns on the medium
3. Environmental Monitoring Test : Exposure of media plates for 4 h as a settle plate or in air sampler or even under laminar air flow may lead reduction in some available moisture on the surface. This may cause development of tiny cracks in the agar or slight shrinkage. This however, does not impact the performance of the media.
4. Each lot of the medium has been tested for the organisms specified on the COA. It is recommended to users to validate the medium for any specific microorganism other than mentioned in the COA based on the user's unique requirement.
5. It is recommended to store the plates at 24-30°C to avoid minimum condensation

## Performance and Evaluation

Performance of the medium is expected when used as per the direction on the label within the expiry period when stored at recommended temperature.

## Methodology

Either streak, inoculate or surface spread the test inoculum (50-100 CFU) aseptically on the plate .

## Quality Control

### Appearance

Sterile Sabouraud Dextrose Agar Plate w/ 1% Glycerol in 90 mm disposable plates.

### Colour of medium

Light amber coloured medium.

### Quantity of medium

30 ml of medium in 90 mm disposable plates

### pH

5.40-5.80

### Does of irradiation (Kgy)

13.00 - 20.00

### Sterility Test

Passes release criteria

### Cultural Response

Growth Promotion was carried out in accordance with the (USP/EP/BP/JP), after an incubation at 20-25 °C for 24-48 hours. Recovery rate is considered as 100% for bacteria growth on Soybean Casein Digest Agar and fungus growth on Sabouraud Dextrose Agar.

### Growth Promotion Test

Growth Promotion was carried out in accordance with the harmonized method of ICH (USP/EP/BP/JP), after an incubation at 30-35 °C for 24-48 hours. Recovery rate is considered as 100% for bacteria growth on Soybean Casein Digest Agar and fungus growth on Sabouraud Dextrose Agar

### Growth Promoting Properties

Growth of microorganism comparable to that previously obtained with previously tested and approved lot of medium occurs at the specified temperature for not more than the shortest period of time specified inoculating  $\geq 100$  cfu (at 30-35°C for 24 hours).

### Indicative properties

Colonies are comparable in appearance and indication reaction to those previously obtained with previously tested and approved lot of medium occurs for the specified temperature for a period of time within the range specified inoculating  $\geq 100$  cfu (at 30-35°C for 24-48 hours).

Organism	Inoculum (CFU)	Growth	Recovery
<i>Candida albicans</i> ATCC 10231 (00054*)	50-100	luxuriant (white colonies)	$\geq 70$ %
<i>Aspergillus brasiliensis</i> ATCC 16404 (00053*)	50-100	luxuriant	$\geq 70$ %



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<i>Saccharomyces cerevisiae</i> ATCC 9763 (00058*)	50-100	luxuriant	>=70 %
<i>Escherichia coli</i> ATCC 8739 (00012*)	50-100	luxuriant	>=70 %
<i>Escherichia coli</i> ATCC 25922 (00013*)	50-100	luxuriant	>=70 %
<i>Escherichia coli</i> NCTC 9002	50-100	luxuriant	>=70 %
<i>Lactobacillus casei</i> ATCC334	50-100	luxuriant	>=70 %
<i>Trichophyton rubrum</i> ATCC 28191		luxuriant	>=70 %

Key : (\*) Corresponding WDCM numbers.

## Storage and Shelf Life

On receipt store between 20-30°C. Use before expiry date on the label. Product performance is best if used within stated expiry period

## Disposal

User must ensure safe disposal by autoclaving and/or incineration of used or unusable preparations of this product. Follow established laboratory procedures in disposing of infectious materials and material that comes into contact with sample must be decontaminated and disposed of in accordance with current laboratory techniques (2,3).

## Further Reading

1. American Public Health Association, Standard Methods for the Examination of Dairy Products, 1978, 14th Ed., Washington D.C.
2. Bacteriological Analytical Manual, 8th Edition, Revision A, 1998. AOAC, Washington D.C.
3. Carlier G. I. M., 1948, Brit. J. Derm. Syph., 60:61.
4. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition.
5. Jorgensen, J.H., Pfaller, M.A., Carroll, K.C., Funke, G., Landry, M.L., Richter, S.S and Warnock, D.W. (2015) Manual of Clinical Microbiology, 11th Edition. Vol. 1.
6. Murray PR, Baron EJ, Tenover JC, Tenover FC (editors) 2003, Manual of clinical Microbiology, 8th ed., ASM, Washington, D.C.
7. Sabouraud K., 1892, Ann. Dermatol. Syphilol, 3:1061.
8. Salfinger Y., and Tortorello M.L. , 2015, Compendium of Methods for the Microbiological Examination of Foods, 5th Ed., American Public Health Association, Washington, D.C.
9. Wehr H. M. and Frank J. H., 2004, Standard Methods for the Microbiological Examination of Dairy Products, 17th Ed., APHA Inc., Washington, D.C.

## Disclaimer

- User must ensure suitability of the product(s) in their application prior to use.
- The product conform solely to the technical information provided in this booklet and to the best of knowledge research and development work carried at CDH is true and accurate
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