

Technical Information

MacConkey Agar RS Plate

Product Code: PM 2702

Application: Recommended for isolating and differentiating Gram negative enteric bacilli from specimens containing swarming strains of Proteus species

Composition**

Ingredients	Gms / Litre		
Peptone	17.000		
Proteose peptone	3.000		
Lactose	10.000		
Bile salts	5.000		
Sodium chloride	5.000		
Neutral red	0.030		
Crystal violet	0.001		
Agar	13.500		
Final pH (at 25°C)	7.1±0.2		

^{**}Formula adjusted, standardized to suit performance parameters

Principle & Interpretation

MacConkey agars are slightly selective and differential plating media mainly used for the detection and isolation of gram- negative organisms from clinical (7,8), dairy (15), food (6,12), water (2), pharmaceutical (4,13) and industrial sources (16). It is also recommended for the selection and recovery of the Enterobacteriaceae and related enteric gram-negativebacilli. USP recommends this medium for use in the performance of Microbial Limit Tests (13).

These agar media are selective since the concentration of bile salts, which inhibit gram-positive microorganisms, is low in comparison with other enteric plating media. The medium M081, which corresponds with, that recommended by APHA canbe used for the direct plating of water samples for coliform bacilli, for the examination of food samples for food poisoning organisms (12) and for the isolation of Salmonella and Shigella species in cheese (15). Other than that this medium is also used for count of coli-aerogenes bacteria in cattle and sheep faeces (7), the count of coli-aerogenes and non-lactose fermenters in poultry carcasses (3), bacterial counts on irradiated canned minced chicken (14) and the recognition of coli- aerogenes bacteria during investigations on the genus Aeromonas (5).

MacConkey Agar is the earliest selective and differential medium for cultivation of enteric microorganisms from a variety of clinical specimens (9,10). The original medium contains protein, bile salts, sodium chloride and two dyes. The selective action of this medium is attributed to crystal violet and bile salts, which are inhibitory to most species of gram-positive bacteria. Gram-negative bacteria usually grow well on the medium and are differentiated by their ability to ferment lactose. Lactose-fermenting strains grow as red or pink colonies and may be surrounded by a zone of acid precipitated bile. The red colour is due to production of acid from lactose, absorption of neutral red and a subsequent colour change of the dye when the pH of medium falls below 6.8. Lactose non-fermenting strains, such as Shigella and Salmonella are colourless, transparent and typically do not alter appearance of the medium.

Peptones are sources of nitrogen and other nutrients. Lactose is a fermentable carbohydrate, bile salts and crystal violet are selective agents that inhibit growth of gram-positive organisms. Neutral red is the pH indicator dye.



Type of specimen

Clinical - faeces, urine and other pathological material, food stuffs and dairy samples, water samples, pharmaceutical samples.

Specimen Collection and Handling

For clinical samples follow appropriate techniques for handling specimens as per established guidelines (8,9).

For food and dairy samples, follow appropriate techniques for sample collection and processing as per guidelines (1,14,17). For water samples, follow appropriate techniques for sample collection, processing as per guidelines and local standards (2). For industrial samples, follow appropriate techniques for sample collection, processing as per guidelines (16).

After use, contaminated materials must be sterilized by autoclaving before discarding.

Warning and Precautions

In Vitro diagnostic use only. Read the label before opening the container. Wear protective gloves/protective clothing/eye protection/face protection. Follow good microbiological lab practices while handling specimens and culture. Standard precautions as per established guidelines should be followed while handling clinical specimens. Safety guidelines may be referred in individual safety data sheets.

Limitations

- 1. Individual organisms differ in their growth requirement and may show variable growth patterns on the medium
- 2. Each lot of the medium has been tested for the organisms specified on the COA. It is recommended to users to validate the medium for any specific microorganism other than mentioned in the COA based on the user's unique requirement.
- 3. It is recommended to store the plates ta 24-30°C to avoid minimum condensation .
- 4. Though the medium is recommended for selective isolation, further biochemical and serological testing must be carried out for further confirmation.
- 5. The surface of the medium should be dry when inoculated.

Performance and Evaluation

Performance of the medium is expected when used as per the direction on the label within the expiry period when stored at recommended temperature.

Methodology

Either streak, inoculate or surface spread the test inoculum (50-100 CFU) aseptically on the plate.

Quality Control

Appearance

Sterile MacConkey Agar RS Plate in 90 mm disposable plate.

Colour

Red with purplish tinge coloured

Quantity of medium

25ml of medium in disposable plate

Reaction

6.90- 7.30



Cultural Response

Cultural response was observed after an incubation at 30-35°C for 18-72 hours. Recovery rate is considered as 100% forbacteria growth on Soybean Casein Digest Agar.

Organism	Inoculum(CFU)	Growth	Recovery	Colour of Colony
Corynebacterium diphtheriae	>=10 ⁴	Inhibited	0%	
type gravis				
Shigella flexneri ATCC12022 (00126*)	50-100	fair to good	30-40%	Colourless
Salmonella Paratyphi AATCC 9150	50-100	Luxuriant	>=50%	Colourless
Salmonella Abony NCTC6017 (00029*)	50-100	Luxuriant	>=50%	Colourless
Proteus vulgaris ATCC13315	50-100	Luxuriant	>=50%	Colourless
Salmonella Typhi ATCC 6539	50-100	Luxuriant	>=50%	Colourless
Staphylococcus epidermidis ATCC 12228 (00036*)	>=10 ⁴	Inhibited	0%	
Escherichia coli ATCC 8739 (00012*)	50-100	Luxuriant	>=50%	Pink red with bile precipitate
Staphylococcus aureus subsp.aureus ATCC 5538 (00032*)	>=10 ⁴	Inhibited	0%	
Salmonella Paratyphi B ATCC 8759	50-100	Luxuriant	>=50%	Colourless
Escherichia coli ATCC25922 (00013*)	50-100	Luxuriant	>=50%	Pink red with bile precipitate
Escherichia coli NCTC 9002	50-100	Luxuriant	>=50%	Pink red with bile precipitate
# Klebsiella aerogenes ATCC 13048 (00175*)	50-100	Luxuriant	>=50%	Pink to red
Salmonella Typhimurium ATCC 14028 (00031*)	50-100	Luxuriant	>=50%	Colourless
Enterococcus faecalis ATCC 29212 (00087*)	50-100	non-poor	<=10%	Colourless to pale pink
Salmonella Enteritidis ATCC 13076 (00030*)	50-100	Luxuriant	>=50%	Colourless
Staphylococcus aureus subsp.aureus ATCC 25923 (00034*)	>=10 ⁴	Inhibited	0%	

^{(*) -} Corresponding WDCM numbers

Storage and Shelf Life

- On receipt store between 20-30°C Use before expiry date on the label.
- Product performance is best if used within stated expiry period.

Disposal

User must ensure safe disposal by autoclaving and/or incineration of used or unusable preparations of this product. Follow established laboratory procedures in disposing of infectious materials and material that comes into contact with sample must be decontaminated and disposed of in accordance with current laboratory techniques (7,8).

[#] Formerly known as *Enterobacter aerogenes*



Further Reading

- American Public Health Association, Standard Methods for the Examination of Dairy Products, 1978, 14th Washington D.C.
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- 5. Eddy B. P., 1960, J. Appl. Bacteriol., 23(2).216-249.
- 6. FDA Bacteriological Analytical Manual, 2005, 18th Ed., AOAC, Washington, D.C.
- 7. Isenberg, H.D. Clinical Microbiology Procedures Handbook 2nd Edition.
- 8. Jorgensen, J.H., Pfaller, M.A., Carroll, K.C., Funke, G., Landry, M.L., Richter, S.S and Warnock., D.W. (2015) Manual of Clinical Microbiology, 11th Edition. Vol. 1.
- 9. MacConkey A., 1905, J. Hyg., 5:333.
- 10. MacConkey A., 1900, The Lancet, ii:20.
- 11. Medrek T. F and Barnes Ella M., 1962, J. Appl. Bacteriol., 25(2),159-168
- 12. Salfinger Y., and Tortorello M.L., 2015, Compendium of Methods for the Microbiological Examination of Foods, 5th Ed., American Public Health Association, Washington, D.C.
- 13. The United States Pharmacopoeia, 2019, The United States Pharmacopeial Convention, Rockville, M.D.
- 14. Thornley Margaret J., 1957, J. Appl. Bacteriol., 20(2), 273-285.
- 15. Wehr H. M. and Frank J. H., 2004, Standard Methods for the Microbiological Examination of Dairy Products 17thEd., APHA Inc., Washington, D.C.
- 16. Williams, (Ed.), 2005, Official Methods of Analysis of the Association of Official Analytical Chemists, 19th Ed., AOAC, Washington, D.C

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