Ready Prepared Media

Technical Information

MiCrome Universal Agar Plate

Product Code: PM 2600

Application: Recommended for for presumptive identification of microorganisms from clinical and non-clinical specimens .

Composition**

Ingredients	Gms / Litre
Peptone	15.000
Chromogenic mixture	2.500
Tryptone	4.000
Agar	13.500
Final pH (at 25°C)	7.2±0.2

**Formula adjusted, standardized to suit performance parameters

Principle & Interpretation

MiCrome Universal Differential Medium is a modification of the medium formulated on basis of the work carried out by Pezzlo (1), Wilkie et al (2), Friedman et al (3), Murray et al (4), Soriano and Ponte (5) and Merlino et al (6). MiCrome Universal Differential Medium is recommended for the presumptive identification of microorganisms from clinical and non- clinical specimens where the medium has broader application as a general nutrient agar for isolation of various microorganisms. This medium helps in the identification of some gram-positive bacteria and gram-negative bacteria on the basis of different colony colours exhibited by them. These colours are formed due to the reactions of genus or species specific enzymes with the two chromogenic substrates incorporated in the medium. *Enterococcus* species, *Escherichia coli* and coliforms produce enzymes which specifically cleave these chromogenic substrates to give characteristically distinctive colony colours. Peptones in the medium serve as sources of amino acids like phenylalanine and tryptophan which aids in indicating tryptophan deaminase activity, thereby facilitating the identification of *Proteus* species, *Morganella* species and *Providencia* species. One of the chromogenic substrate is cleaved by ß-glucosidase enzyme possessed by Enterococci resulting in the formation of bluish green colonies. *Escherichia coli* possesses the enzyme ß- galactosidase which specifically cleaves the other chromogenic substrate resulting in the formation of purple coloured colonies. *Escherichia coli* can be differentiated and confirmed from other similar coloured colonies, by performing the indole test.

Coliforms cleave both the chromogenic substrates forming blue to purple coloured colonies. Colonies of *Proteus, Morganella* and *Providencia* species appear brown due to tryptophan deaminase activity. Peptone and tryptone provide nitrogenous, carbonaceous compounds, essential growth nutrients and also serve as a source of amino acids.

Type of specimen

Clinical samples : urine, faeces, Food and dairy samples, Water samples.

Specimen Collection and Handling

For clinical samples follow appropriate techniques for handling specimens as per established guidelines (7,8).
For food and dairy samples, follow appropriate techniques for sample collection and processing as per guidelines (9,10,11). For water samples, follow appropriate techniques for sample collection, processing as per guidelines and local standards (12).After use, contaminated materials must be sterilized by autoclaving before discarding.

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Warning and Precautions

In Vitro diagnostic Use. For professional use only. Read the label before opening the pack. Wear protective gloves/protective clothing/eye protection/ face protection. Follow good microbiological lab practices while handling specimens and culture. Standard precautions as per established guidelines should be followed while handling clinical specimens. Safety guidelinesmay be referred in individual safety data sheets.

Limitations

1. Since it is an enzyme-substrate based reaction, the intensity of colour may vary with isolates.
2. Individual organisms differ in their growth requirement and may show variable growth patterns on the medium
3. Each lot of the medium has been tested for the organisms specified on the COA. It is recommended to users to validate the medium for any specific microorganism other than mentioned in the COA based on the user's unique requirement.

Performance and Evaluation

Performance of the medium is expected when used as per the direction on the label within the expiry period when stored at recommended temperature.

Methodology

Either streak, inoculate or surface spread the test inoculum (50-100 CFU) aseptically on the plate.

Quality Control

Appearance

Sterile HiCrome™ Universal Agar, in 90mm disposable plates with smooth surface and absence of black particles/cracks/ bubbles.

Colour of medium

Light amber coloured

Quantity of medium

25ml of medium in disposable plate

pH

7.00-7.40

Sterility check

Passes release criteria

Cultural Response

Cultural characteristics observed after an incubation at 35-37°C for 18-48 hours.

Organism	Inoculum(CFU)	Growth	Recovery	Colour of Colony
Enterococcus faecalis ATCC 29212 (00087*)	50-100	Luxuriant	>=70%	blue,small
Escherichia coli ATCC 25922 (00013*)	50-100	Luxuriant	>=70%	purple
Klebsiella pneumoniae ATCC 13883 (00097*)	50-100	Luxuriant	>=70%	blue-green mucoid
Pseudomonas aeruginosa ATCC 27853 (00025*)	50-100	Luxuriant	>=70%	colourless (greenish pigment may be observed)

Proteus mirabilis ATCC12453	50-100	Luxuriant	>=70%	light brown
Staphylococcus aureus subsp. aureus ATCC 25923 (00034*)	50-100	Luxuriant	>=70%	golden yellow
Salmonella Typhi ATCC 6539	50-100	Luxuriant	>=70%	colourless
Salmonella Typhimurium ATCC 14028 (00031*)	50-100	Luxuriant	>=70%	colourless

(*) - Corresponding WDCM numbers

Storage and Shelf Life

- On receipt store between 2-8°C Use before expiry date on the label.
- Product performance is best if used within stated expiry period.

Disposal

User must ensure safe disposal by autoclaving and/or incineration of used or unusable preparations of this product. Follow established laboratory procedures in disposing of infectious materials and material that comes into contact with sample must be decontaminated and disposed of in accordance with current laboratory techniques (7,8).

Further Reading

1. Pezzlo M (1998), Clinical Microbiology Reviews 1:268-280
2. Wilkie M.E.,Almond M.K.,Marsh F.P.(1992),British Medical Journal 305:1137-1141.
3. Friedman M.P. et al (1991), Journal of Clinical Microbiology, 29:2385-2389.
4. Murray P., Traynor P. Hopson D., (1992), Journal of Clinical Microbiology 30:1600-1601.
5. Soriano F., Ponte C., (1992), Journal of Clinical Microbiology 30:3033-3034.
6. Merlino et al (1995) Abstr. Austr. Microbiol. 16(4):17-3.
7. Isenberg, H.D. Clinical Microbiology Procedures Handbook 2nd Edition.
8. Jorgensen, J.H., Pfaller, M.A., Carroll, K.C., Funke, G., Landry, M.L., Richter, S.S and Warnock., D.W.(2015) Manual of Clinical Microbiology, 11th Edition. Vol. 1.
9. American Public Health Association, Standard Methods for the Examination of Dairy Products, 1978, 14th Ed., Washington D.C.
10. Salfinger Y., and Tortorello M.L. 2015, Compendium of Methods for the Microbiological Examination of Foods, 5th Ed., American Public Health Association, Washington, D.C.
11. Wehr H. M. and Frank J. H., 2004, Standard Methods for the Microbiological Examination of Dairy Products, 17th Ed., APHA Inc., Washington, D.C.
12. Lipps WC, Braun-Howland EB, Baxter TE,eds. Standard methods for the Examination of Water and Wastewater,24th ed. Washington DC:APHA Press; 2023.

Disclaimer :

- User must ensure suitability of the product(s) in their application prior to use.
- The product conform solely to the technical information provided in this booklet and to the best of knowledge research and development work carried at **CDH** is true and accurate
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