

## Technical Information

### Motility Test Agar

**Product Code : SM1260**

**Application:-** Recommended for detection of bacterial motility.

#### Composition\*\*

Ingredients	Gms / Litre
Tryptose	10.000
Sodium chloride	5.000
Agar	5.000
Final pH (at 25°C)	7.2±0.2

\*\* Formula adjusted, standardized to suit performance parameters.

#### Principle & Interpretation

Bacterial motility can be observed directly on microscopic slide or it can be visualized on motility media having agar concentration of 0.4% or less (4). Use of such semisolid media to observe or detect motility was reported by Tittsler and Sandholzer (6). Motility Test Medium is a modification of their formulation. Motility can be visualized as a diffused zone of growth flaring out from the line of inoculation (4). Hanging-drop technique in motility tests has practical difficulties, which is efficiently eliminated by use of culture-based methods using semi-solid media, as in semisolid media; the results obtained are macroscopic and cumulative.

Tryptose serve as a source of essential growth nutrients required for bacterial metabolism. Sodium chloride maintains the osmotic equilibrium of the medium. Small amount of agar helps to create a semisolid medium. Bacterial motility can be observed directly by examination of the tubes following incubation. Inoculation is done by stabbing through the centre of the medium. Incubate at appropriate temperature for 18-40 hours. Non-motile organisms grow only along the line of inoculation whereas motile organisms grow away from the line of inoculation or may show growth even throughout the medium. All weak or equivocal motility results should be confirmed by flagellum stain or by direct wet microscopy (hanging drop) (1,5).

#### Type of specimen

Isolated microorganism

#### Specimen Collection and Handling:

With inoculating needle, stab centre of medium to approximately one-half of depth.(4) After use, contaminated materials must be sterilized by autoclaving before discarding.

#### Warning and Precautions :

Read the label before opening the container. Wear protective gloves/protective clothing/eye protection/ face protection. Follow good microbiological lab practices while handling specimens and culture. Standard precautions as per established guidelines should be followed while handling specimens. Safety guidelines may be referred in individual safety data sheets.

#### Limitations :

- 1.Growth from an 18-24 hr pure culture should be used. (5)
2. All weak or equivocal motility results should be confirmed by flagellum stain or by direct wet microscopy (hanging drop) (1,5).

#### Performance and Evaluation

Performance of the medium is expected when used as per the direction on the label within the expiry period when stored at recommended temperature

#### Methodology

Motility Test Agar is a ready to use solid media in glass bottle.The medium is pre-sterilized, hence it does not need sterilization. Medium in the bottle can be melted either by using a pre-heated water bath or any other method. Slightly loosen the cap before melting. When complete melting of medium is observed dispense the medium in tubes as butts /slants or in plates as desired and allow to solidify. If on plate, either streak, inoculate or surface spread the test inoculum (50-100 CFU) aseptically.

## Quality Control

### Appearance

Sterile Motility Test Agar in glass bottle.

### Colour medium

Light yellow coloured

### Quantity of medium

100 ml of medium in glass bottle.

### Reaction

7.00-7.40

### Sterility Test

Passes release criteria

### Cultural Response

Cultural characteristics observed after an incubation at 35 - 37°C for 18 - 48 hours

Organism	Inoculum (CFU)	Growth	Motility
<i>Escherichia coli</i> ATCC 25922 (00013*)	50-100	luxuriant	positive, growth away from stabline causing turbidity
# <i>Klebsiella aerogenes</i> ATCC 13048 (00175*)	50-100	luxuriant	positive, growth away from stabline causing turbidity
<i>Klebsiella pneumonia</i> ATCC 13883 (00097*)	50-100	luxuriant	negative, growth along the stabline, surrounding medium remains clear
<i>Salmonella</i> Enteritidis ATCC 13076 (00030*)	50-100	luxuriant	positive growth away from stabline causing turbidity
<i>Staphylococcus aureus</i> subsp. <i>aureus</i> ATCC 25923 (00034*)	50-100	luxuriant	negative, growth along the stabline, surrounding medium remains clear

Key : (\*) Corresponding WDCM numbers.

(#) Formerly known as *Enterobacter aerogenes*

## Storage and Shelf Life

On receipt store between 15-25°C Use before expiry date on the label. Product performance is best if used within stated expiry period.

## Disposal

User must ensure safe disposal by autoclaving and/or incineration of used or unusable preparations of this product. Follow established laboratory procedures in disposing of infectious materials and material that comes into contact with sample must be decontaminated and disposed of in accordance with current laboratory techniques (2,3).

## Further Reading

1. DAmato R. F., and Tomfohre K. M., 1981, J. Clin. Microbiol., 14 (3), 347-348.
2. Isenberg, H.D. Clinical Microbiology Procedures Handbook 2<sup>nd</sup> Edition.
3. Jorgensen, J.H., Pfaller, M.A., Carroll, K.C., Funke, G., Landry, M.L., Richter, S.S and Warnock., D.W. (2015) Manual of Clinical Microbiology, 11th Edition. Vol. 1.
4. Koneman E. W., Allen S. D., Janda W. M., Schreckenberger P. C., Winn W. C. Jr., (Eds.), 1992, Colour Atlas and Textbook of Diagnostic Microbiology, 4th Ed., J. B. Lippincott Company.
5. MacFaddin J. F., 1985, Media for Isolation-Cultivation-Identification-Maintenance of Medical Bacteria, Vol. 1, Williams and Wilkins, Baltimore
6. Tittsler R. P. and Sandholzer L. A., 1936, J. Bacteriol., 31:575.



Dehydrated Culture Media  
Bases / Media Supplements

## Disclaimer :

- User must ensure suitability of the product(s) in their application prior to use.
- The product conform solely to the technical information provided in this booklet and to the best of knowledge research and development
- work carried at **CDH** is true and accurate
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